Recent advances in **data structures** for storing **sets of k-mer sets**

Rayan Chikhi Institut Pasteur

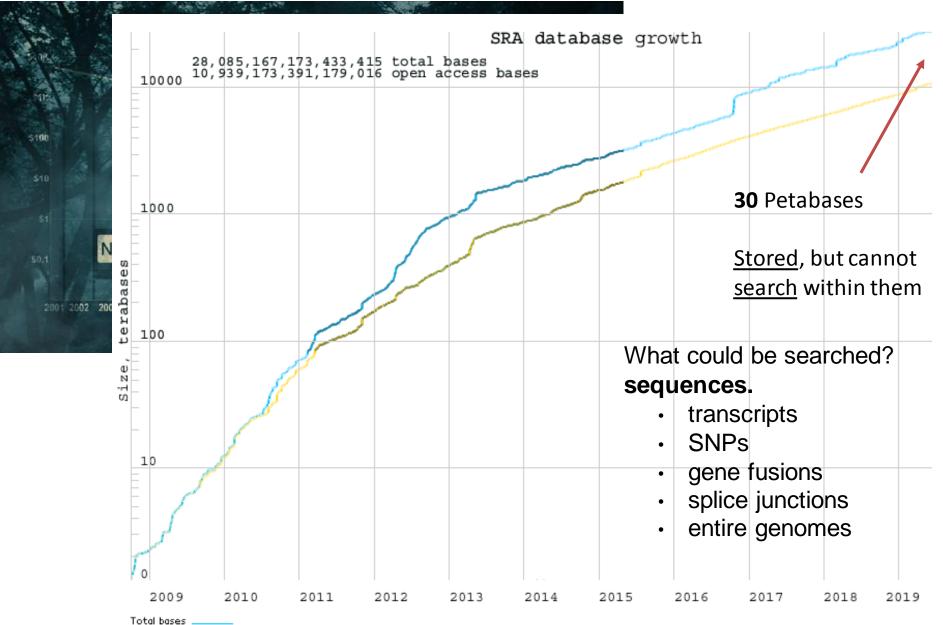
CGSI 2019

Searching huge databases of sequences

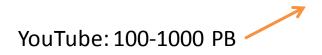
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CGSI 2019

https://www.ncbi.nlm.nih.gov/sra/docs/sragrowth/



Open access hases



NCBI SRA database : 30 PB



Institut Pasteur: 8 PB



Your laptop: 0.001 PB



SRA	SRA •		Search
		Advanced	Help



SRA

Sequence Read Archive (SRA) makes biological sequence data available to the research community to enhance reproducibility and allow for new discoveries by comparing data sets. The SRA stores raw sequencing data and alignment information from high-throughput sequencing platforms, including Roche 454 GS System®, Illumina Genome Analyzer®, Applied Biosystems SOLiD System®, Helicos Heliscope®, Complete Genomics®, and Pacific Biosciences SMRT®.

Search results

Items: 1 to 20 of 19964	NextSeq 500 paired end sequencing (ERR3407135)
	Metadata Analysis (alpha) Reads Download
 NextSeq 500 paire 1 ILLUMINA (Illumina Accession: ERX34307 	Filtered Download What does it do? What can the filter be applied to?
NextSeq 500 paire	< 1 1 346553 > View: Sological reads technical reads
 2. 1 ILLUMINA (Illumina Accession: ERX34307) NextSeq 500 paire 3. 1 ILLUMINA (Illumina Accession: ERX34307) 	1. ERR3407135.1 ERS3549882 name: NB551234:144:HL523AFXY:1:11101:5421: member: default >gnl SRA ERR3407135.1.1 NB551234:144:HL523AFXY:1:11101:5421:1076 <i>F</i> (Biological) ACCTGAGCGCGCAGCTCCAGCTAAATCAAACGCGGCGCGGAATTTGGGATGTTCCATCAGT TTCCAGGCGCGCTGTTGCCCTCACGTCACAGTAACTGAAGCTGCCAAATATCACGG GTAAGCGTGGTAAGGCGTTTCGGGATCGCCA * >gnl SRA ERR3407135.1.2 NB551234:144:HL523AFXY:1:11101:5421:1076 <i>R</i> (Biological) ACCAGACAGCGGGGAATACCACCTCTTCCAGCCGTTGTTTCCAACCAA

Ν	шн	U.S. National Library of Medicine NCBI Na	Iational Center for Biotechnology Information
	BLAS	ST [®] » blastn suite	
			Sequence Read Archive Nucleotide BLAST
	blastn]	
	Ent	er Query Sequence	BLASTN programs search SRA databases using a nucleotide query. 😡
	Enter	accession number(s), gi(s), or FASTA sequence(s	e(s) 😡 <u>Clear</u> Query subrange 😡
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			study, or submission), title, the scientific name or tax id. Only 20 top suggestions will be shown.

What could be gained by large-scale sequence searches?

- conditions that express a given novel isoform
- gene fusions across TCGA
- metagenomic samples containing bacterial strain
- Anti-microbial resistance:
 - detection of drug-resistant genes
 - phylogeny of plasmids carrying AMR genes



https://m.photofunia.com/

Problem definition

- Given many FASTQ files (=*experiments*):
 - Experiment 1
 - Experiment 100,000
- And a sequence s
- Enumerate all the experiment(s) where s appears
- e.g. "ACAGTATGGTTGGGGGAAAAG" -> Experiments {23 (human RNAseq), 1523 (human RNAseq), 82499 (human gut metagenome)}

Searching within sequences, techniques

Solution 0: grep

Solution 1: build a *huge* dictionary

Solution 2: FM-index

Solution 3 and others: settle for k-mer searches

Solution 0: grep

How fast is grep?

- SRA has .fastq.gz files
- gunzip: 60 MB/sec



pugz: ~400 MB/sec [Kerbiriou, Chikhi 2019]

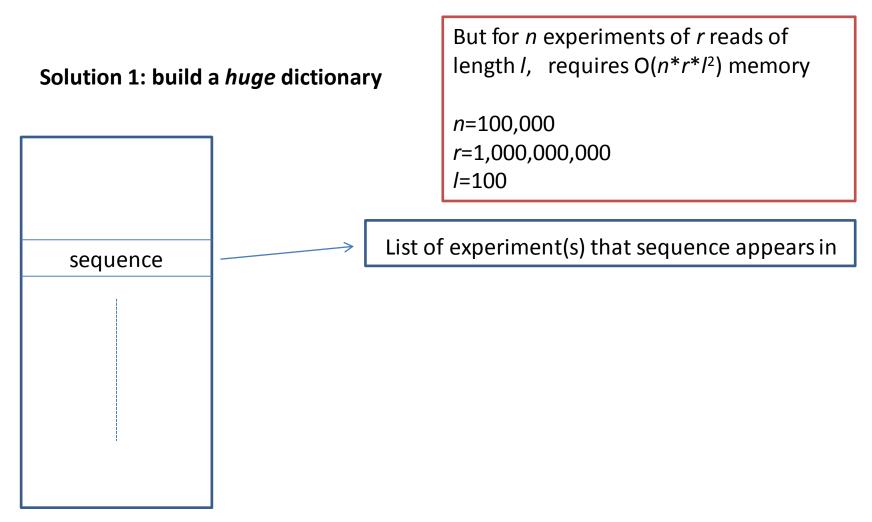
Time:

- 60 GB Illumina human FASTQ: 16 minutes, 1 thread
- 1,000 of them:

~1 day, 10 threads

• SRA (5M files):

~1 year, 100 threads



Solution 1: build a *huge* dictionary

What if we indexed only small sequences of fixed length?

How many in, say, all RefSeq proteobacteria (E. coli & al)?

21,883 genomes, 97 GB of FASTAs -> 31 billion sequences [Kerbiriou, personal comm, k=31]

Lower bound of ~ 85 GB to represent [Conway, Bromage 2012; Chikhi et al, 2014] (as opposed to 31 billion times 31 nucleotides)

That's only for a small subset of *genomes*, not even FASTQs...

Solution 2: FM-index

(2017, Genome Research)

Method-

Using reference-free compressed data structures to analyze sequencing reads from thousands of human genomes

Dirk D. Dolle,^{1,6} Zhicheng Liu,^{1,2,6} Matthew Cotten,¹ Jared T. Simpson,^{3,4} Zamin Iqbal,⁵ Richard Durbin,¹ Shane A. McCarthy,¹ and Thomas M. Keane^{1,2}

- Built a BWT of 2,705 human Illumina WGS error-corrected reads
- Size: ~500 GB + ~5 TB metadata (origin of reads)
- 1 k-mer search ~= 10 ms

k-mer search

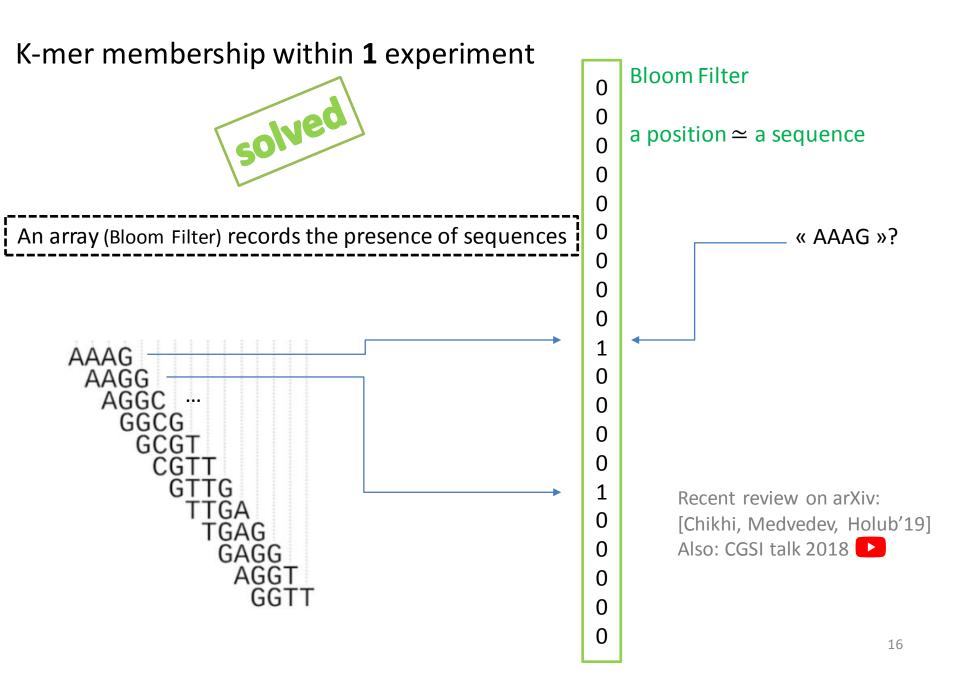
• k-mer Sequence of fixed length k

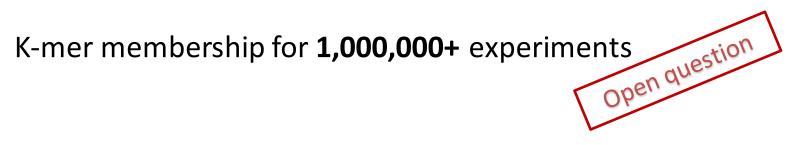
ref: ACGATGACATGAT 4-mers: ACGA CGAT GATG ATGA

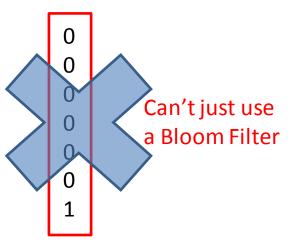
...

- Membership query
 -> Is x in S?
- Enables more complex queries

arbitrary sequence s <-> decomposition of s into k-mers e.g. SNP <-> set of ~k k-mers







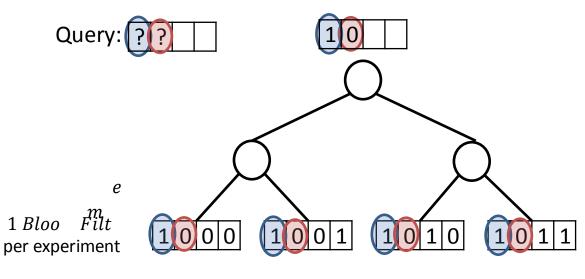
Why:

- *Single* SRA Bloom Filter
- 1 BF per experiment

- : ~ petabytes, and pooled
- : ~ hour-long query (150ms * 5M exps)

Sequence Bloom Trees

[Solomon & Kingsford 2016]



Slide: P. Medvedev

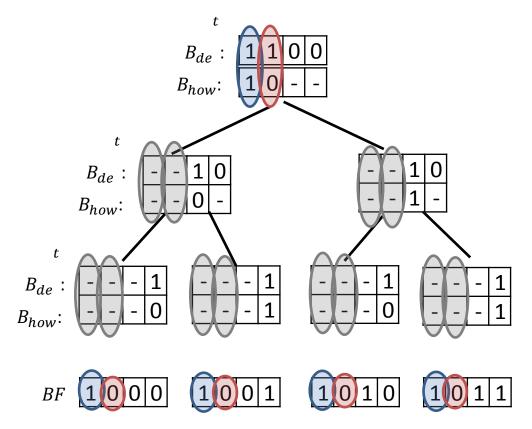
Fast search through pruning

- blue seq matches ALL experiments
- query can stop at root

• AllSome Sequence Bloom Trees [Sun, Harris, Chikhi, Medvedev 2017]

HowDeSBT

[Harris, Medvedev 2019]



Slide: P. Medvedev

SBT performance in a nutshell

Data	Size on disk
2,000 raw experiments	~15,000 GB
AllSome SBT (2017)	142 GB
HowDe SBT (2019)	14 GB

- Smaller space than raw data
- Resides fully on disk
- 1 search = 5 seconds

BIGSI

[Bradley, .., Iqbal 2019]

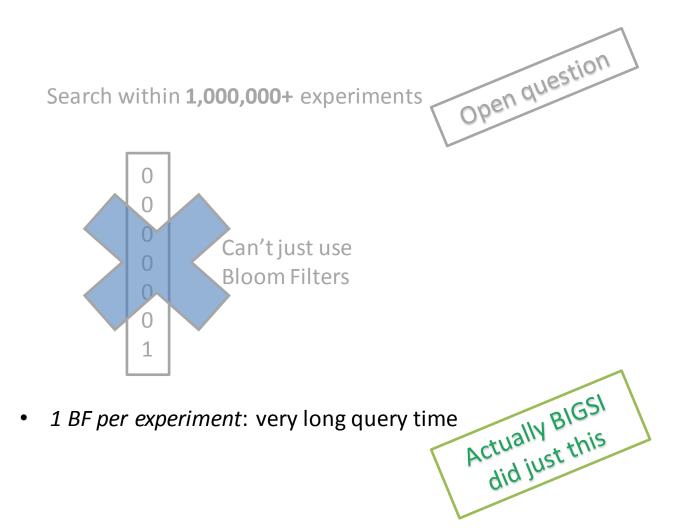
FEBRUARY 2019 VOL 37 NO 2 WW.nature.com/naturebit technology

> Ultrafast searching for microbial sequences Virus capture in complex metagenomics samples RNA editing with endogenous enzymes

CCA

G

Recall, a few slides ago...



BIGSI is a vector of Bloom filters

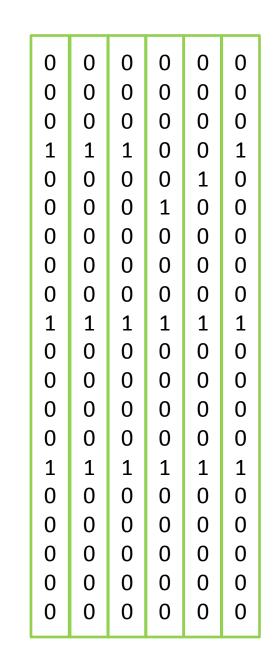
[Bradley et al, 2019]

(or equivalently, a matrix of bits)

- Rows = k-mers
 Columns = experiments
- All the BFs have the <u>same size</u> How is this possible? *Microbial data*
- 1 search = 0.3 second



Also the technique behind



Many other methods

- SBT: Solomon and Kingsford 2016 Nature biotechno.
- AllSomeSBT: Sun et al. 2018 RECOMB
- SSBT: Solomon and Kingsford 2018 RECOMB
- HowDeSBT: Harris and Medvedev 2019 RECOMB-Seq
- BIGSI: Bradley et al. 2019 Nature biotechno.
- COBS: Bingman et al. 2019 *arXiv*
- Cortex: Iqbal et al. 2012 Nature
- BFT: Holley et al. 2016 AMB
- VARI: Muggli et al. 2017 Bioinformatics
- VARI-Merge: 2019 accepted to ISMB
- Rainbowfish: Almodaresi et al. 2017 WABI
- Mantis: Pandey et al. 2018 Cell
- Mantis+MST: Almodaresi et al. 2019 RECOMB
- SeqOthello: Yu et al. 2018 Genome Biology
- Metannot: Mustafa et al. 2018
- Multi-BRWT: Karasikov et al. 2018

Slide: C. Marchet

Review: [Marchet *et al*, in preparation '19] Short overview in [Chikhi, Medvedev, Holub'19]

Conclusion

- Sets of k-mer sets, a powerful representation for sequencing data
- Booming area since 2016
- No method is *really* user-friendly yet
- Many similar experiments: SBT *et al*
- Many experiments of same size: BIGSI et al
- And many others I didn't talk about
- Versatile method:

[???]

A Tera increase in sequencing production in the past 25 years						
Genes & Operons	1990	Kilo = 1,000				
Bacterial genomes	1995	Mega = 1,000,000				
Human genome	2000	Giga = 1,000,000,000				
Human microbiome	2005	Tera = 1,000,000,000,000				
50K Microbiomes	2015	Peta = 1,000,000,000,000,000				
what is expected for the next 15 years ? (a Giga?)						
200K Microbiomes	2020	Exa = 1,000,000,000,000,000				
1M Microbiomes	2025	Zetta = 1,000,000,000,000,000,000,000				
Earth Microbiome	2030	Yotta = 1,000,000,000,000,000,000,000				

Source: @kyrpides

Thank you for your attention! Any questions?

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