Space-efficient and exact de Bruijn graph representation based on a Bloom filter

Rayan Chikhi, Guillaume Rizk

ENS Cachan Brittany / IRISA, France AlgoRizk, France

WABI 2012





TRANSITION FROM THE PREVIOUS TALK

The **previous talk** dealed with a **succinct de Bruijn graph representation**. **This talk** covers exactly the **same topic**, with some differences :

- (Hopefully) simpler data structure
- Less succinct
- Implemented in an assembly program

OUTLINE

Presentation of the data structure

Analysis

Assembly aspects

Results

Perspectives

Presentation of the data structure

de Bruijn graph

Nodes are *k*-mers, edges are (k - 1)-overlaps between nodes.

 $\mathsf{GAT} \longrightarrow \mathsf{ATT} \longrightarrow \mathsf{TTA} \longrightarrow \mathsf{TAC} \longrightarrow \mathsf{ACA} \longrightarrow \mathsf{CAA}$

Only **nodes** need to be encoded, as **edges** are inferred.

How to encode the de Bruijn graph using as little space as possible?

Memory usage

- Explicit list : $2k \cdot n$ bits
- Self-information of n nodes :

$$\log_2\left(\binom{4^k}{n}\right)$$
 bits

20 bits per node.

(illustration for k = 25)

50 bits per node

[Conway, Bromage 11]

[Idury, Waterman 95]

Bloom filter

Bit array to represent any set with a "precision" of ϵ .

• a proportion ϵ of elements will be wrongly included (*false positives*).

To represent a set of *n* elements, requires $\approx 1.44 \log_2(\frac{1}{\epsilon}) \cdot n$ bits.

Storing *k*-mers in a Bloom filter :



Bloom filter

Bit array to represent any set with a "precision" of ϵ .

• a proportion ϵ of elements will be wrongly included (*false positives*).

To represent a set of *n* elements, requires $\approx 1.44 \log_2(\frac{1}{\epsilon}) \cdot n$ bits.

Storing *k*-mers in a Bloom filter :



Bloom filter

Bit array to represent any set with a "precision" of ϵ .

• a proportion ϵ of elements will be wrongly included (*false positives*).

To represent a set of *n* elements, requires $\approx 1.44 \log_2(\frac{1}{\epsilon}) \cdot n$ bits.

Storing *k*-mers in a Bloom filter :



AAA (hash value 0) present? **Yes**, maybe : either a true or a false positive.

Set of **nodes** : {TAT, ATC, CGC, CTA, CCG, TCC, GCT} Graph as stored in a Bloom filter :

[Pell et al 12]



Black nodes : true positives ; Red nodes : false positives

Insight : to **traverse** the graph from **true positive** nodes, only a **small fraction of the false positives** need to be **avoided** (*critical false positives*, *CFP*).



Proposed method

Store **nodes** on **disk** for sequential enumeration, and in **memory** store the **Bloom filter** + the critical FPs **explicitly**.



Analysis

Construction time (for *n* nodes)

Assume that *k*-mer arithmetic takes constant time.

- ▶ Bloom filter construction : *O*(*n*)
- cFP construction :
 - Enumeration of neighbors of all graph nodes, keeping only Bloom-positive neighbors : O(n)
 - Intersection between Bloom-positive neighbors and nodes, with limited memory usage : O(k log(k) n)

OPTIMAL BLOOM FILTER SIZE

Structure size per k-mer, k=27



Size of the Bloom filter (bits / k-mer)

Dependence on the parameter k

Optimal structure size per k-mer



k-mer size

Result statement

The de Bruijn graph can be encoded using

$$\underbrace{1.44 \log_2(\frac{16k}{2.08})}_{\text{Bloom}} + \underbrace{2.08}_{\text{cFP}}$$

bits of memory per node.

k = 25: **13** bits per node.

- Below the self-information (20 bits/node for k = 25)
- The part stored in memory doesn't support enumeration of nodes, only traversal

Graph-based assemblers typically **modify the graph** to remove artifacts (variants, errors).

Is it possible to perform *de novo* assembly with this (immutable) structure? \rightarrow Yes, using localized traversal. [RC DL, WABI 11]

Assembly aspects

Traverse the graph greedily, according to these rules :







Traverse the graph greedily, according to these rules :



Example : Start with an empty graph

Traverse the graph greedily, according to these rules :



Example : Pick a new node, construct the first portion



Traverse the graph greedily, according to these rules :



Example : Construct the second portion



Traverse the graph greedily, according to these rules :





ASSEMBLER IMPLEMENTATION

Need to determine the set of solid nodes (seen *k*-mer counting $\geq x$ times) Current methods (e.g. Jellyfish) require more memory than our structure We designed a constant-memory k-mer counting procedure Graph traversal Nodes which have already been traversed need to be marked No extra information can be stored in our structure We used a separate hash table to remember if branching or dead-end nodes have already been visited. Contigs construction Consensus from each path obtained by localized traversal

Results

USEFULNESS OF CFP STRUCTURE



Assembly : E. coli, k = 23

COMPLETE *de novo* HUMAN GENOME ASSEMBLY

N50 : length 1 at which half of the assembly contains sequences of length $\geq l$

Human genome assembly	Minia	C. & B.	ABySS	SOAPdenovo
Contig N50 (bp)	1156	250	870	886
Sum (Gbp)	2.09	1.72	2.10	2.08
> 95% Accuracy (%)	94.6	-	94.2	-
Nb of nodes/cores	1/1	1/8	21/168	1/40
Time (wall-clock, h)	23	50	15	33
Memory (sum of nodes, GB)	5.7	32	336	140

ROUGH PERFORMANCE COMPARISON WITH OTHER HUMAN GENOME ASSEMBLIES



Perspectives

PERSPECTIVES

Applications

Why assemble a human genome again?

To exhibit novel structural variations

[Iqbal 11]

As a **benchmark**, for the immense number of (meta)genomes that will be sequenced next

Future of sequencing

Predictions :

DNA assembly Relevant until 10-100 kbp high-accuracy read lengths

RNA assembly, **metagenomics** and **metatranscriptomics** No announced technology other than **Illumina** permits high depth of sampling.

 \rightarrow My opinion is that **short-read assembly** (with paired reads) will remain a hot topic for a few years.

PERSPECTIVES

Potential applications of Minia codebase :

- Huge metagenomic assemblies
- ► Transcriptome assembly
- Alternative splicing detection
- SNP detection

n (KisSplice module replacement) (KisSnp 2, with R. Uricaru & P. Peterlongo)

(with Genoscope) (Inchworm replacement)

- Structural variants detection
- Gap-filling of scaffolds
- Read compression

AVAILABILITY

Manuscript available at **minia.genouest.org**. To obtain the source code of Minia (pending license) :

Now Email me Next month Website above Acknowledgements : Dominique Lavenier, GenScale team (IRISA, France)

Thank you! Any questions?